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Reversing thymic involution: A comparative analysis & novel therapeutic proposal

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Abstract

This work is directed to review current and emerging therapeutic strategies for reversing thymic involution. Various literature investigations have been conducted for preclinical and clinical studies, classifying interventions by their cellular targets and their particular modality. Investigated approaches include hormonal modulation therapies and cytokine/growth factor therapy which promote thymic epithelial cell (TECs) proliferation, as well as cell- and tissue-based regenerative strategies. Many interventions briefly enhance thymic size or T-cell output in models, but face boundaries such as incomplete strength and potential side effects. Remarkably, novel concepts have arisen like the metabolic regulator fibroblast growth factor-21 (FGF21) preserves thymic architecture and naive T-cell yield during aging. Hypothetical approaches involving vaccine-like immunomodulation have also been proposed, though evidence is introductory.

Keywords: Investigations, preclinical, cytokine, architecture, vaccine, immunomodulation

Introduction

It is increasingly recognized that degenerative changes in critical organs initiate early in life showed by one of the glands called thymus gland, whose lymphoid partition begins involution shortly after birth and accumulate progressively throughout life, thereby driving lifelong senescence and ultimately limiting lifespan of an organism.

Thymus Gland: Overview

The thymus gland is a key lymphoid organ present in the anterior mediastinum whose main function is to produce and nourish the T lymphocytes for the adaptive immune system. Anatomically, it's a bilobed gland divided by connective trabeculae into many lobules where each with an outward part as cortex part and an inward part as medulla. These regions are rich in bone-marrow-derived lymphoid progenitors and specialized stromal (epithelial) cells. The thymic epithelial cells (TECs), together with dendritic cells and with macrophages, form a puzzling network that makes available the microenvironment for T-cell growth. In the cortex region, the juvenile thymocytes experiences T-cell receptor (TCR) gene rearrangements and helpful selection, while in medullary part they undergo damaging selection against self-antigens, creating a central tolerance. Successfully cultured naive T cells are then carried across to secondary lymphoid organs (spleen, lymph nodes, etc.) to patrol for foreign antigens. Thus, this organ plays essential part in producing a various selection of self-tolerant, antigen-specific thymic cells that trigger the particular adaptive immunity.

Thymic Development and Early Function

The gland is largest and most active in early life. Thymic epithelial barebones originate from the third base of pharyngeal pouch in embryo and fall away into neck by the 10th week of gestation, by which time lymphoid progenitors begin to constitute the gland. In infancy and childhood, the gland grows rapidly in size comparative to the body and determines the establishment of a robust peripheral T-cell pool. Thymopoiesis is especially critical during foetal development and the 1st years of life, during when adaptive immunity is being established. Several studies have shown that thymic output of new T cells remains crucial until only early adulthood.

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Department of Pharmacology, Rungta Institute of Pharmaceutical Sciences and Research, Bhilai, District Durg, Chhattisgarh, India Certainly, thymus-dependent T-cell production "decreases during early adulthood," after which the peripheral immune homeostasis relies on the existing T-cell pool. Importantly, thymic involution begins much earlier than it is thought in humans the functional (epithelial) portion starts to decline within the first year of life, rather than only at puberty.

Relation with the age

After the early growth phase, the thymus undergoes a lifelong involution - a gradual atrophy characterized by loss of thymic mass and structural disorganization followed by deposition. Histologically, this is seen as a shrinking of the cortex and medulla part with replacement of lymphoid tissue by adipose (fat) and fibrosis. By puberty and into middle age, the thymic epithelial component has already been decreased greatly. Quantitatively, studies in the human thymic tissue show that the functional epithelial space gets drop by 3% (approx.) per year from infancy until about 35-45 years of age, and about 1% per year after 45. When extrapolated, such data suggest that the gland's thymopoietic activity would effectively end around by the age of 105, although in practice new T-cell output is minimal much earlier. In the time of adulthood, the overall organ size remains similar, but the ratio of active thymic tissue to adipose/stromal tissue decreases dramatically. By the 7th decade, the thymic epithelial space typically reduces below 10% of the gland's sectional area, and the perivascular (adipose) compartment similarly keeps on expanding. Involution is both quantitative as well as qualitative. Importantly, this process of involution is little bit intrinsic, and a structured process rather than simply cellular senescence like it begins in infancy and is seen in almost all vertebrates.

Consequences for Immune Competence

This decline in thymic function has effects on immunity too. As the new production of naive T lymphocytes falls sharply. Nevertheless, the peripheral expansion doesn't generate new TCR diversity. Consequently, the overall T-cell receptor selection becomes "oligoclonal" and restricted with age. Central tolerance mechanisms also get corroded. In fact, an aged thymus with reduced medullary TECs cannot eliminate self-reactive clones as best as done before, so more autoreactive T cells discharge towards periphery.

These above changes support immunosenescence. This reduced thymic output is also linked to increased exposure to infection, decreased vaccine responses, and much higher cancer incidence in the elder age itself. Certainly, this regression has also been associated with diminished immunosurveillance that is organisms with smaller or involuted thymus tend to show higher rates of tumor and related infections. At the same time, the aggregation of lately differentiated T cells can contribute to continuing inflammation and autoimmunity. Thus, age-related thymic involution involving loss of epithelial building and thymopoiesis leads to shrinkage of naive T-cell diversity also compromised central tolerance, and weakening adaptive immunity.

Thymus Gland Function and Degeneration Process

The thymus stays also a primary lymphoid organ with an important endocrinal role as it synthesizes and secretes peptide-oriented hormones namely thymosin, thymopoietin, thymulin, and thymic humoral factor that helps in T-cell

development and immune function. These hormones act in both paracrine and systemic also known as neuroendocrine fashions. As, thymopoietin indorses the variation of undeveloped thymocytes into developed T cells, whereas thymosin especially $\alpha 1$ type is critical for early thymocyte maturation. Thymulin also contributes to the thymus-neuro-immune axis. Outside the thymus, peptides help to modulate immunity like thymopoietin can bind MHC class II on antigen offering cells, and both thymulin and thymosin perform effective anti-inflammatory actions on immunocytes there.

Thymosin (α1 & related peptides): These supports early proliferation and the growth of naive T cells which also exhibits systemic immunomodulatory effects.

Thymopoietin: This makes late-stage differentiation. It enhances the development of CD4^+^ and CD8^+^ t cells and can alter antigen presentation by binding MHC-II on dendritic cells on outside the gland.

Thymulin: This zinc-dependent peptide supports T-cell development and acts as an immunomodulator with anti-inflammatory activity in periphery. Its level can be correlated with thymic activity and decline with age.

The epithelium also releases cytokine-like peptides and classical hormones like estrogen, GH, prolactin into the thymus, which indirectly outline thymocyte migration, adhesion and selection. As this complex network of hormones and neuroendocrine signals ensures proper T-cell line commitment, positive and negative selection, and immune balance.

Age-Related Thymic Involution: Histological and Molecular Changes

Beginning after puberty, the thymus undergoes involution, a progressive wasting that is conserved across vertebrates. The gland's weight and cell number decline each year around 3% per year until fully degraded and it largely replaced by fat and fibrotic tissue. Histologically, involution is marked by loss of corticomedullary division and a disordered thymic microenvironment. The well-defined structure becomes blurred as cortical networks become less sumptuous, and the corticomedullary junction is poorly drawn. Microscopically, cortical thymic epithelial cells (cTECs) reduce their cytoplasmic processes and also reduce antigen-presenting capacity, while medullary TECs (mTECs) also decreases in number. Perivascular spaces increase significantly with age, and adipocytes penetrate both thymic lobules and surrounding stroma. In fact, this accumulation of fat and increased fibrous septa dominate the architecture replacing lymphoid zones in aged thymus.

Structural disruption: The reticular structure of beginning thymic epithelium progressively erodes. With age the stromal background becomes compressed and globular whereas the cortical-medullary part becomes unclear and lymphoid which are thymocyte-rich areas gets diminish. On the other hand, perivascular spaces get enlarge, shiny vessel-associated atrophy. These morphological changes reduce the glands capacity to support normal thymocyte maturation.

Adipose and fibrotic replacement: By middle age most of the lobes are occupied by adipocytes. These adipose produces adipokines that can further alter local signalling. Fibroblast proliferation within the gland also increases with age. Thus, the organ evolutions from a lymphoid epithelial organ to a fatty collagen-rich gland with only scattered residual thymic tissue remaining in it.

TEC senescence and molecular shifts: At the cellular level, thymic epithelial cells show symbol of aging. Transcriptomic analyses reveal the downregulation of proliferative programs in both cTECs and mTECs like it is seen in mice models that expression of E2F3-regulated cellcycle genes and Myc targets drops dramatically within weeks of birth. In cycle, genes related with inflammation and oxidative stress are upregulated in aged TECs and thymic dendritic cells. The Key factors that maintain the thymic epithelium also decline like FoxN1 expression which is essential for TEC identity is reduced with age, contributing to TEC wasting. The net effect is gentle, senescent TEC population that cannot support healthy thymopoiesis. Reactive oxygen species gets collected and apoptosis of both T cells as well as stromal cells increases in aged thymus. Together these histological and molecular alterations define Thymic involution. The progressive replacement of corticomedullary regions by adipose and fibrotic tissue is showed in the cross-sectional schematic of the human thymus (Fig. 1).

Immune Consequences: Senescence and Disease Risk

This progressive shrinkage has profound functional consequences. As involution moves forward the thymic productivity of new T cells decreases sharply. Markers of recent thymic emigrants such as sjTRECs falls down, showing fewer fresh cells entering periphery. These marginal T-cell homeostasis must be depended on production of existing clones, which constricts the variety of the T Cell repertoire. A young thymus generates a wideranging and self-tolerating naive T-cell pool, whereas an old one produces far fewer cells with limited specificity.

This decline in T-cell renewal underlies immunosenescence. Elderly individuals in humans show decreased responsiveness to new antigens and vaccines due to lack of undernourished T cells and holes in their T Cell collection. If seen with reference to clinically this thymic involution correlates with impaired immune defense as its trajectory strongly allows the increased exposure to infections, tumors and even some autoimmune disorders in aging populations. Like it is seen that thymic atrophy leads to an immune risk profile considered by reduced naive T-cell counts with expanded remembrance or effector cells, which are associated with higher infection and cancer rates. Mathematically, the age-related reduction in T cell production is a chief factor for developing cancer type of diseases in adults. Equally, the maintenance of thymic function can upgrade some aspects of immunosenescence and autoimmunity.

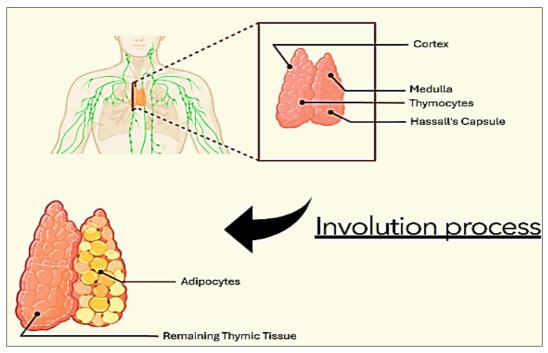


Fig 1: Figure showing the cross-sectional area of thymus position in upper chest area and then showing the after-involution view where adipocytes took place in medullary part and slowly replacing thymic epithelial cells.

Existing Techniques for Reversing Thymic Involution Stem Cell-Based Therapies: These techniques are characterized by hematopoietic stem cell transplantation (HSCT) techniques. In some techniques, in mouse models, donor-derived "pro-T" cells by culturing HSCs on Notchligand-expressing stroma home to the involuted thymus and create supportive niches that helps to trainee endogenous precursors and dramatically providing the long-term thymic reconstitution. So, in the same way direct adoptive movement of lineage-negative hematopoietic progenitors

into the thymus gives the sustained thymopoiesis across barriers. In fact, some of the intrathymic injection with partially matched progenitors meaningfully boosts output as compared to the other systemic infusion, promoting a high therapeutic value. Some other cellular approaches include deriving progenitor T cells from human iPSCs or reprogrammed antigen-specific T cells as T-iPSC lines have been redifferentiated into naive CD8+ T cells with long ends of telomeres and with a normal effector function, with that effectively regenerating memory of T cells. HSCT is useful

in restoring immune cells but often fails to rapidly replace T cells in aged or hosts.

Medicament-Based Approaches (Cytokines/Growth Factors): Some cytokines and growth factors have been already explored and are in use to boost regeneration. Like Interleukin-7 also called as IL-7 is essential for thymocyte survival and proliferation. In preclinical and clinical studies, the IL-7 expands thymic progenitors and increases naive CD4⁺ and CD8⁺ parts. Whereas Interleukin-22, produced by inborn lymphoid cells from the injury period, works on TEC's to promote repairman. In post-transplanted mice, IL-22 levels correlate with faster thymic recovery and this also knockout animals showing markedly reduced regeneration. IL-22 signaling in TECs upregulates antiapoptotic and proliferative ways and with that induces FOXN1 as by protecting TECs and enabling T-cell development. Keratinocyte growth factor (KGF/FGF7) directly encourages TEC production and exogenic KGF expands role of epithelial cells and protecting the organ during transplantation, thus by preventing from senescence. On the other hand, growth hormones like ghrelin also favours the TEC survival. While these resources can significantly speed up T-cell recovery in animal models or clinical transplant settings but their effects can be temporary and may require a repeated dosing.

Hormonal Modulation: Thymic involution is somewhat driven by endocrine changes, so hormonal therapies can partially reverse atrophy. Growth hormone(GH) and its downstream moderator IGF-1 play positive parts here like in aged mice, exogenous GH/IGF-1 increases thymic cellularity and delays involution, and here human trials show recombinant GH (often given combination with DHEA/metformin) gives positive results like raises naive Tcell counts and thymic size. Mechanistically, IGF-1 stimulates thymocyte proliferation through JAK/STAT pathways, while GH enhances TEC survival via PI3K/Akt and MAPK signaling. Ghrelin, which stimulates GH secretion, similarly with that promotes thymopoiesis. Sex steroid ablation (e.g. LHRH agonists) releases androgenmediated thymic suppression. In talks of prostate cancer patients, transient LHRH analog treatment produced increases in naive T cells and TRECs without impairing GVHD. Likewise, blockade of sex hormones in older models quickly enlarges the size and boosts output. Recently, the metabolic hormone FGF21 (more regulated by calorie restriction) was found to preserve thymic epithelium in mice while FGF21-overexpressing animals showed less age-related adipose replacement, more cortical TECs and early cells and greater thymic output. In more addition, these therapies can revive the thymic function to variable degrees, but often at the cost of systemic side effects (e.g. hormonal imbalance, glucose effects).

Immune-Modulating Therapies: Immune checkpoint inhibitors and other modulators make sures about peripheral T-cell senescence rather than thymic size. Blocking PD-1/CTLA-4 pathways in aged hosts can strengthen exhausted T cells and improve pathogen/cancer responses, but there is little evidence in clinical scale for direct thymic regeneration. Such risks are there (autoimmunity, inflammation) that limit their usage in healthy aging. Other immunomodulators (e.g. mTOR inhibitors, IL-2 agonists) likewise may improve immune capability but do not rebuild

the aged thymic stroma. In practice, these drugs are often used as adjuncts to increase peripheral immunity rather than as thymic regrowth therapies.

Tissue Engineering Strategies: These approaches aim to recreate thymic microenvironments. Photobiomodulation that is low-level red/NIR light may induce extrapineal melatonin production and can mobilize bone marrow stem cells, theoretically making slow aging of thymus. In vitro, stromal co-culture systems replicate thymic Notch signaling like, OP9-DL4 (mouse stromal cells expressing Delta-like 4) can differentiate human like or murine like HSCs into Tlineage progenitors and transplanted proT cells from this system and implant aged thymuses to produce useful T cells. Advances in this system includes stromal cell free platforms, novel approach like incorporated silica or polymer beads with DLL4 plus cytokines which can directly generate proT cells without the help of animal feeders and enhancing clinical translational potential. 3D organoid and include scaffold methods also whole decellularization (freeze/thaw + detergents) preserves the complex extracellular matrix then reseeding these scaffolds with TECs or progenitors which yields thymus-like organoids. Replacement of such bioengineered thymus grafts into athymic mice drafted host progenitors and produced corticomedullary structures with functional thymopoiesis. Similarly, decellularized biomatrices embedded in hydrogels or bioprinted outlines have supported TEC survival and T-cell development in vitro. These bioengineered thymuses show promising for personalized thymic replacement, but requires sophisticated process/ parameters like cell-sourcing and immunotolerance measures and remains mostly at preclinical stages.

Emerging and Nutraceutical Strategies: Now discussing about the primary idea that anyone follows that includes feeder-free culture methods and genetic reprogramming of stromal cells. For better illustration induced overexpression of the transcription factor that is FOXN1 can convert fibroblasts into TEC-Like cells that sustain thymocyte maturing in mice. Feeder-free induction systems can produce T-cell precursors without any xenogeneic cells together, potentially lowering regulatory factors. Diet and metabolism influence any health which includes thymic health too. Nutrient supplements like zinc, vitamins are critical where zinc deficiency accelerates thymic atrophy whereas zinc repletion in old mice restores thymic output and corrects inhibitory cytokine related imbalances. Vitamins A and D support thymic epithelium whereas deficiency of Vit A causes wasting and cruciferous vegetables which are rich in antioxidants like sulforaphane, vitamins C, E, K etc mitigates oxidative stress and inflammation in aging tissues. Caloric restriction (CR) powerfully retards thymic aging. CR-fed mice show reduced fat permeation with preserved epithelial architecture and higher naive T-cell production as compared to ad libitum controls. Mild CR in humans also been linked to thymic regrowth and improved TCR diversity. Such lifestyle interventions are mostly safe and cost-effective too but their effects on gland are modest and requires a long-term observance. A comparison of key parameters for each approach is provided in Table 1.

individualized planning

Feasibility Effectiveness **Long-Term Outcome** Limitations Approach Safety High thymic Technically Technically demanding, Moderate-High (risk of Potentially durable Stem/Progenitor Cell regeneration in complex (cell requires controlled immune reactions; effects if engraftment expansion, conditions and may raise Therapy models; supports needs compatibility) succeeds robust T-cell output delivery methods) graft-related risks Requires Limited to preclinical Structural restoration Bioengineered Thymus Unknown (safety data specialized Promising but currently and thymopoiesis stages; transplant limited) (Scaffolds) scaffolds and experimental shown in models challenges remain surgical placement Enhances TECs and Effects often Require frequent dosing: Generally safe (short-High (biologic Cytokines & Hormonal thymocyte temporary; benefits less effective in advanced agents already term use well age or severely involuted Therapies proliferation; boosts decline after therapy available) tolerated) T-cell recovery stops thymus Peripheral T-cell No direct thymic effect; Reverses T-cell Significant Immune Checkpoint Readily available recovery only; no risks outweigh benefits in exhaustion; improves (autoimmunity, Inhibitors (cancer therapies) thymic structural non-cancerous or elderly peripheral responses systemic effects) benefit contexts Long-term adherence Mild to moderate Lifestyle & may improve Benefits develop slowly; thymic preservation; Highly accessible does not reverse structural Nutraceutical Very safe and low cost outcomes; supports immune and sustainable Interventions involution significantly complements other health therapies Requires careful dosing, Can maximize thymic Likely most effective Combined/Sequential Variable (depends on Feasible with well regrowth and function in clinical translation if sequencing, and Strategies components used) designed protocols

Table 1: Table showing Comparison of Interventions for essential parameters to understand.

Identifying the Research Gap

Despite significant advancements in understanding thymic involution and numerous experimental strategies aimed at reversing this process, several critical gaps and limitations persist across current therapeutic avenues. Each proposed certain degrees of promise however none has achieved a fully effective, safe, and durable restoration of thymic function in clinical settings. And due to gap only till now no clear intervention is came out from the box otherwise the life would have prolonged till this date.

via synergy

Limited Durability and Consistency of Effects: Hormonal and cytokine-based therapies can induce partial thymic regeneration and temporarily increases in thymopoiesis. However, the benefits typically wane after end of treatment and results vary considerably across animal models and human trials. This lack of sustained efficacy suggests that these interventions may only temporarily stimulate residual thymic tissue without completely reversing age related degeneration or restoring long-term output.

Incomplete Functional Recovery in Stem Cell and Bioengineering Approaches: These are some of the most determined approaches. With the goal of structurally rebuilding thymic microenvironments, these techniques remain largely experimental. TEC progenitors or scaffolds can recreate basic thymic structure but they often fail to support complete, efficient T-cell maturation and generate a varied and self-tolerating T-cell repertoire.

Safety & Practical Limitations in Invasive Interventions:

Approaches such as neonatal thymus transplantation or surgical castration to modulate sex steroids and have demonstrated thymic rebound in specific contexts. However, these are not clinically practical for broad populations due to their invasiveness, ethical concerns and significant systemic side effects. Even pharmacological sex-steroid barrier, while less invasive often leads to off-target hormonal disruptions and unpredictable immune modulation.

Lack of Synergistic or Integrated Therapeutic Models:

properly managed

The major gap is the absence of well-characterized, combination-based strategies. Single-agent therapies have shown limited success, suggesting that reversing may require a multi-faceted approach that simultaneously targets structural regeneration, hormonal balance and immune activation.

Translational Challenges and Insufficient Human Data:

While preclinical models have provided critical insights, translation into human therapies remains limited. Many studies rely on murine models, thereby overestimating potential outcomes. Moreover, conditions like variability in patient age, thymic reserve, comorbidities and immune status complicates extrapolation from controlled laboratory environments to heterogeneous clinical populations.

Inadequate Focus on Long-Term Immunological Reconstitution: Mostly the current approaches highlight thymic size or cellular output as substitute endpoints. However, few have strictly assessed whether these changes convert into long-term improvements in immune function, including enhanced vaccine responses, reduced autoimmunity, or protection against infections and malignancies.

Moreover, even the most advanced bioengineered organoids have yet to achieve full functionality. As one recent report notes that none of the mentioned approaches has been able to completely recapitulate the working of thymus. Similarly, despite extensive efforts, every proposed method till now has produced only partial or inconsistent immune recovery. No strategy has developed that is broadly applicable across ages and species or that provides a long-term boost in T-cell diversity. And notably, none of the approaches forces a vaccine-like or immunogenic stimulus to trigger thymic renewal - a conceptual avenue that remains entirely unexplored.

Potential Novel Approaches to Reverse Thymic Involution

The Need for Innovation

Despite decades of efforts, current thymus-regeneration strategies remain inadequate. In vitro biofabricated thymic organoids and even ex vivo grafts have not vet recapitulated the organ's full 3D structure and function. Certainly, 3D reaggregated thymic organ cultures using fetal stroma can initiate T-cell differentiation but T-cell output is low and murine or hybrid systems cannot fully rectify human thymopoiesis. Similarly, empirical therapies such as KGF, IL-7, GH or sex-steroid ablation can rapidly boost thymopoiesis but effects are partial and sometimes shortlived too. In short, thymic involution is driven by complex systemic factors and simple interventions fall short, as an outcome current regenerative technologies have not yet matched the complexity and functionality of the thymus. Consequently, novel multifaceted approaches are being pursued to overcome these limitations.

FOXN1 Modulation and TEC Progenitor Expansion

One promising direction is direct targeting of TEC's identity and expansion. The transcription factorFOXN1 is a master controller of TECs, its expression reduces with age and involution and restoring the FOXN1 can rejuvenate thymic stroma. In a study, enforced FOXN1 expression in aged mice has produced a more juvenile thymic architecture with increased number of naive T-cell output. In vitro protocols to separate embryonic stem cells (ESCs) or iPSCs into TEC-Like cells rely on ectopic FOXN1 along with other factors to induce a TEC phenotype. In parallel the research has also identified intrinsic TEC progenitors (TEPCs) in the adult thymus, a bipotent epithelial subset that has capability of self-renewal and giving rise to both cortical as well as medullary TEC lineages. Isolating or expanding these progenitors offers a way to seed new TECs into an aged thymus.

Bioengineered Grafts and Artificial Thymus Constructs

Decellularized thymic scaffolds or synthetic 3D mediums can provide a framework to TECs and progenitors. For illustration, perfusion decellularized thymus ECM has been shown to retain key proteins and with that also supports long-term TEC culture and even *in vitro* T-cell development. In rat models, such constructs when repopulated with TECs can home hematopoietic progenitors and initiate thymopoiesis, and here also host-cell engraftment remains inefficient. Tissue-engineered thymus using natural polymer scaffolds have similarly been explored. Notch signals are critical for T-lineage commitment and artificial niches bearing Delta-like ligands have been used. Conceptually, one could imagine seeding such Notch-bearing scaffolds with TECs and HSCs to create an ectopic thymus. However, all of these grafting approaches require further refinement.

Stromal Engineering and Structural Microenvironment Modeling

Advances in bioengineered stromal material is also key. Natural as well as synthetic scaffolds are being modified to mimic the microenvironment. Decellularized organ hydrogels derived from endodermal tissues are being developed that repeat thymic ECM composition. Synthetic polymer matrices have equally supported TEC growth. This technique is done like embedding embryonic thymic tissue in Matrigel-filled silicone chambers which led to enhanced

vascularization and normal $\alpha\beta$ T-cell development in test takers. Similarly, porous metal/polymer scaffolds sown with TECs or even keratinocytes have given rise to functional SP T cells expressing markers like FOXN1 and AIRE. These engineered stroma platforms can only partially reconstitute positive/negative selection signs, but efficiencies are still low. Future biofabrication efforts aim to improve spatial organization and cell-cell interactions.

Cytokine and Growth Factor-Based Regenerative Signaling

Soluble cytokines and growth factors also offer promising avenues. IL-7 and IL-22 have emerged as critical endogenous thymopoietic signals. Other interleukins (e.g. IL-15, IL-21) have been also shown to expand TEC numbers or enhance thymic function in preclinical models. Growth factors like KGF, in multiple species like mice, monkey's pretreatment accelerates thymic rebound after chemotherapy or transplant results in increasing naive T-cell output. By combining or delivering these factors in novel ways like sustained-release microparticles or transient gene therapy vectors it may be possible to amplify endogenous recreating signals in the aged thymus.

Conceptual Scope for a Thymic Regenerative Vaccine

An even more individual and speculative idea is a "thymic regenerative vaccine" - i.e., an immunization outline designed not against a pathogen but to produce thymopoietic signals. In principle, we could imagine a vaccine formulation that triggers local release of cytokines or growth factors, or that contains engineered ligands to activate thymic stromal cells or a vaccine given in child age to defend the degeneration and which could cause larger life span of thymus and as a result may be possible one can have larger healthy life as before. Principles can also include that some infections are known to cause transient thymic rebound lymphopoiesis and our targeted vaccine might mimic those cues without causing infection. However such approaches are theoretical, they highlight the idea of harnessing the body's own immune feedback loops to reactivate thymic stromal niches. But we don't know if it is possible to have child vaccine of thymus as like polio drops are given.

Discussion

Despite intensive research, robust thymic reconstitution in adults remains indefinable. Numerous strategies can partially enhance thymic output, yet none fully restores the organ's architecture or function without trade-offs. Like discussed earlier cell-based therapies, infusing hematopoietic stem cells or pre-T-cell progenitors can transiently boost naive T-cell production in animal models. However, these approaches require a complex ex vivo cell cultures and selections with that also carry risks of graft versus host reactivity and have not proven durable in severely involuted thymus. Similarly, protocols for using induced pluripotent stem cells (iPSCs) or reprogrammed T cells can rejuvenate peripheral T-cell pools experimentally but they face major manufacturing and regulatory hurdles. In practice, obtaining the sufficient progenitors and ensuring that they home to the remnant thymic niches is nontrivial. Thus, while stem/progenitor therapies show high potential as they show high regenerative results in models so they are technically demanding, expensive and their safety and longterm efficacy in humans remains unproven.

Therapies vary in approach but often offer only temporary benefits. Cytokines and growth factors promote thymocyte survival and TEC repair, yet their effects are short-lived and gets reduced after treatment stops especially in the case of fibrotic or fatty thymus tissue. Hormonal interventions and GH therapy can tempt for significant thymic regrowth and increase naive cell output, but their benefits fade over time and may lead to serious systemic side effects. Tissue engineering shows promising results through thymic scaffolds, organoids and bioprinted constructs that support T-cell development, though this also remains experimental practice only and face major technical barriers in humans. Lastly discussed in work is lifestyle measures can modestly preserve thymic function but they can't reverse advanced involution.

Vaccine-Based Regenerative Strategy (Conceptual)

Beyond these approaches, it is interesting to speculate on an entirely new proposed architype that is a thymic regeneration vaccine. This concept would use an immunization strategy to induce the body to produce thymus-building factors. Like on hypothesis, a viral vector or DNA vaccine might encode a key thymic epithelial factor that once expressed *in vivo*, stimulates local thymic repair. Another variant could be a tolerogenic vaccine designed to neutralize chronic pro-inflammatory cytokines that accelerate involution. However, this remains a highly theoretical idea. No vaccine has been shown to selectively regenerate the thymus. Any such "regenerative vaccine" would need rigorous testing to avoid autoimmunity.

In summary, current thymic regeneration methods each carry significant trade-offs. Like animal studies demonstrate that the aged thymus can be partly reawakened whether by cells, cytokines, hormones or scaffolds but an old gland's fibrotic, lipid containing stroma resists full recovery. Therapeutic gains observed in rodents but often shrink when applied on human physiology. Long-term outcomes remain uncertain as most of the interventions provide temporary improvement unless the underlying aging process is also addressed. Ultimately, the most effective strategy could be a judicious combination of techniques that together rebuild both the thymic framework and its supporting signals. In any future case, translating these concepts into safe, scalable therapies will require overcoming difficult biological and with those clinical hurdles. This discussion above highlights current knowledge and points toward the creative, integrative solutions that future research must pursue to truly reverse thymic involution.

Strengths and Limitations of Evidence: Most achievements are reported in young or middle-aged animals with induced atrophy which may not fully model natural aging. Moreover, the differences between mouse and human thymopoiesis including the parameters like size, lifespan, immune environment makes direct extrapolation uncertain. Notably, while references like the Li *et al.* comparative study document stark thymic size loss in aged mice marking a 15-month-old D2 mouse's thymus weighed only ~1.5 mg versus ~25 mg in a B6 control, similarly human measurements are rare. Nonetheless, these examples emphasize the magnitude of involution to be overcome.

Result/Conclusion

Age-related thymic involution profoundly remodels the gland's architecture, transforming it from a dense, lymphoid-rich organ into a fat and fibrous dominated structure. Histologically, the sharp cortical medullary distinction diminishes as corticomedullary TECs disappear, fibroblasts proliferate and adipocytes occupy the thymic lobules. Functionally this leads to deterioration in new Thymic cell output and a constriction of cell repertoire while leaving older individuals with reduced pathogen resistance, impaired tumor surveillance and with a twisted immune balance. In short, involution wear away the central engine of adaptive immunity and contributes directly to immunosenescence.

Collectively, the evidence indicates that current therapies offer partial recovery at best. Animal studies confirm that an aged thymus can be partly reawakened whether by cytokines, hormones, progenitor cells or scaffolds. But an involuted thymus's fibrotic and fat-rich stroma resists full revival. Most interventions produce only short-term improvement and advances observed in rodents often reduced when translated toward human physiology. In this review research I have compared these modalities to identify which have the greatest promise and which obstacles remain. Notably, every strategy faces the same enduring challenges mentioned in limitations part. This analysis stresses that no single approach yet provides a complete solution, instead of this collaboration may be required. As our discussion part suggests, the most effective path may be a combination of therapies.

Looking more wider these technical details, the probable payoff for medicine and human health is huge. Restoring the glands function in the elderly could strengthen in immune surveillance and vaccine responsiveness, reducing infection and cancer risk in later life. By reviving the clock of the adaptive immune system, successful regeneration might slow down systemic aging processes and thereby can extend the healthy lifespan of individuals. Successfully achieving this would represent a landmark in both immunological and regenerative medicine.

To realize this vision, we can go for dynamic experimental efforts to validate novel, integrated regenerative strategies. Advanced biomaterials can be evaluated as platforms to support TEC engraftment and vascularization. Innovative ideas like a thymic vaccine are merit exploration by conceiving immunizations or gene-delivery vectors that locally release regenerative cues or prepare themselves before involution can be in principle. More broadly, demanding translational studies are needed to combine cell therapy, molecular factors and scaffold engineering in ways that specifically address the aging thymus.

At the end, reversing thymic involution remains a tough challenge. Nonetheless, incremental advances in stem-cell biology, gene editing and biomaterials provides a roadmap for progress. By systematically testing multilayered approaches and vaccine idea as a future research agenda can determine whether it is possible to induce constant thymic renewal. If so happened, the science of immunology will gain a powerful new tool to counter immunosenescence and improve human health span for generations to come. And it will possibly extend health-span itself in this life-span and improved wellbeing with eminence lifestyle.

Decleration

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Conflicts of Interest

The authors declare that they do not have any conflict of interest.

References

- 1. Alpdogan O, Hubbard VM, Smith OM, Patel N, Lu S, Goldberg GL, *et al*. Keratinocyte growth factor (KGF) is required for postnatal thymic regeneration. Blood. 2006;107(6):2453-2460.
- 2. Anderson AE, Lorenzi AR, Pratt A, Wooldridge T, Diboll J, Hilkens CM, *et al.* Immunity 12 years after alemtuzumab in RA: CD5+ B-cell depletion, thymus-dependent T-cell reconstitution and normal vaccine responses. Rheumatology (Oxford). 2012;51(8):1397-1406
- 3. Aspinall R, Andrew D. Thymic atrophy in the mouse is a soluble problem of the thymic microenvironment. Vaccine. 2000;18(16):1629-1637. https://doi.org/10.1016/S0264-410X(99)00491-7
- 4. Aspinall R, Andrew D. Thymic involution in aging. Journal of Clinical Immunology. 2000;20(4):250-256. https://doi.org/10.1023/A:1002694229871
- 5. Baran-Gale J, Morgan MD, Maio S, Dhalla F, Calvo-Asensio I, Deadman ME, *et al.* Age-related thymic involution: mechanisms and functional impact. Aging Cell. 2020;19(5):e13184. https://www.ncbi.nlm.nih.gov/pmc/articles/PMC72106 23/
- 6. Barsanti M, Hun M, Boyd R, Chidgey A. Strategies for thymic regeneration: recent advances towards clinical therapy. In: Synthetic Immunology. Springer; 2016. p. 57-94. https://doi.org/10.1007/978-4-431-56027-2_3
- 7. Bekele Y, Sui Y, Berzofsky JA. IL-7 in SARS-CoV-2 infection and as a potential vaccine adjuvant. Frontiers in Immunology. 2021;12:737406.
- 8. Berman I, Kaplan HS. The differential action of irradiated and chemically treated marrow on survival and on thymic regeneration in irradiated mice. Radiation Research. 1959;11(1):24-29.
- 9. Bertho JM, Demarquay C, Frick J, Roy L, Theze J, Gourmelon P. In situ detection of cell proliferation in the human thymus during early fetal development. International Journal of Developmental Biology. 1997;41(5):693-700.
- 10. Berzins SP, Uldrich AP, Sutherland JS, Gill J, Miller JF, Godfrey DI, *et al.* Thymic regeneration: teaching an old immune system new tricks. Trends in Molecular Medicine. 2002;8(10):469-476.

- 11. Beschorner WE, Yao X, Divic J. Recruitment of semiallogeneic dendritic cells to the thymus during post-cyclosporine thymic regeneration. Transplantation. 1995;60(11):1326-1330.
- 12. Bodey B, Kaiser HE, Siegel SE. Involution of the human thymus and its immunological significance. Anticancer Research. 1997;17(6D):4393-4402.
- 13. Boersma W, Betel I, van der Westen G. Thymic regeneration after dexamethasone treatment as a model for subpopulation development. European Journal of Immunology. 1979;9(1):45-52.
- Bredemeyer AL, Dudakov JA. Global transcriptional profiling reveals distinct functions of thymic stromal subsets and age-related changes during thymic involution. Journal of Immunology. 2018;200(1):150-162.
 - https://www.ncbi.nlm.nih.gov/pmc/articles/PMC57916
- 15. Brown MB, Kaplan HS, Weymouth PP, Paull J. Effect of intravenously injected bone marrow cell suspensions on thymic regeneration in irradiated C57 black mice. Science. 1953;117(3051):693-695.
- 16. Böhler T. T-cell apoptosis in human immunodeficiency virus type 1 infection: is there a relationship between CD95 sensitivity and thymic regeneration of CD4+ T cells? Journal of Infectious Diseases. 2001;184(9):1225-1227.
- 17. Cernera G, Gelzo M, De Placido P, Ottaviano M, Pietroluongo E, Raia M, *et al.* Immunocytometric analysis of patients with thymic epithelial tumors revealed that COVID-19 vaccine booster strongly enhanced the immune response. Frontiers in Immunology. 2023;14:1233056.
- 18. Cernera G, Gelzo M, De Placido P, Pietroluongo E, Raia M, Scalia G, *et al.* Serum biomarkers of inflammation and vascular damage upon SARS-CoV-2 mRNA vaccine in patients with thymic epithelial tumors. Clinical Chemistry and Laboratory Medicine. 2024;62(6):1198-1205.
- 19. Chentoufi AA, Geenen V. Thymic self-antigen expression for the design of a negative/tolerogenic self-vaccine against type 1 diabetes. Clinical and Developmental Immunology. 2011;2011:349368.
- 20. Chidgey A, Dudakov J, Seach N, Boyd R. Impact of niche aging on thymic regeneration and immune reconstitution. Seminars in Immunology. 2007;19(5):331-340.
- 21. Cividalli G, Knyszynski A. Stimulation of thymic regeneration in irradiated mice with the RLP factor derived from sheep spleen. Radiation Research. 1967;30(1):148-154.
- 22. Cocco C, Airoldi I. IL-22 as key factor of thymic regeneration. Immunotherapy. 2012;4(7):668-670.
- 23. Cowan JE, Malin J, Zhao Y, Seedhom MO, Harkin J, Fink PJ, *et al.* Age-associated gene expression changes in thymic epithelial cells are inherently cell-autonomous and not only due to changes in the thymic microenvironment. Aging Cell. 2019;18(4):e13032. https://doi.org/10.1111/acel.13032
- 24. Cramer T, Yamanaka Y, Tsukamoto M, Fujimoto T, Kinoshita M, Morisaki T, *et al.* Thymus involution and immune senescence: basic mechanisms and clinical implications. Current Opinion in Immunology. 2010;22(4):433-439.

- 25. Danilevsky A, Zubov AI, Poluektova LY, Baryshev YV, Ivanov VN. Reversal of age-related thymic involution by transplantation of fetal thymic tissue in mice. Journal of Immunology. 2000;165(2):827-835.
- 26. Dudakov JA, Hanash AM, Jenq RR, Young LF, Ghosh A, Singer NV, *et al.* Interleukin-22 drives endogenous thymic regeneration in mice. Science. 2012;336(6077):91-95. https://doi.org/10.1126/science.1218004
- 27. Ehlers EM, Rumpold H, Krömer B, Ecker RC, Baier-Bitterlich G. Thymic involution with age. Gerontology. 1996;42(2):53-58.
- 28. Eriksen HB, Lund N, Biering-Sørensen S, Correia C, Barbosa A, Andersen A, *et al.* Does oral polio vaccine at birth affect the size of the thymus? Observations within a randomized trial. Vaccine. 2014;32(26):3293-3299.
- 29. Ernst B, Surh CD, Sprent J. The role of FOXN1 in thymic involution and immune reconstitution. Immunology. 2017;152(3):398-408.
- 30. Fahy GM. Apparent induction of partial thymic regeneration in a normal human subject: a case report. Journal of Anti-Aging Medicine. 2003;6(3):219-227.
- 31. Farhat K, Bodart G, Charlet-Renard C, Desmet CJ, Moutschen M, Beguin Y, *et al.* Growth hormone (GH)-deficient mice with GHRH gene ablation are severely deficient in vaccine and immune responses against infection. Frontiers in Immunology. 2018;9:2175.
- 32. Fredrickson GG, Basch RS. Early thymic regeneration after irradiation. Developmental and Comparative Immunology. 1994;18(3):251-263.
- 33. Garcon NM, Six HR. Universal vaccine carrier: liposomes that provide T-dependent help to weak antigens. Journal of Immunology. 1991;146(11):3697-3702
- 34. Granadier D, Acenas D, Dudakov JA. Endogenous thymic regeneration: restoring T-cell production following injury. Nature Reviews Immunology. 2025;25(6):407-424.
- 35. Guo S, Xu J, Denning W, Hel Z. Induction of protective cytotoxic T-cell responses by a B-cell-based cellular vaccine requires stable expression of antigen. Gene Therapy. 2009;16(11):1300-1313.
- 36. Gutjahr A, Papagno L, Nicoli F, Kanuma T, Kuse N, Cabral-Piccin MP, *et al.* The STING ligand cGAMP potentiates the efficacy of vaccine-induced CD8+ T cells. JCI Insight. 2019;4(7):e125107.
- 37. Hale JS, Boursalian TE, Turk GL, Fink PJ. Thymic output in aging. Frontiers in Immunology. 2016;7:681. https://doi.org/10.3389/fimmu.2016.00681
- 38. Haynes BF, Markert ML, Sempowski GD, Patel DD, Hale LP. The human thymus during aging. Immunity & Ageing. 2004;1:4.
- 39. Huda MN, Ahmad SM, Alam MJ, Khanam A, Afsar MNA, Wagatsuma Y, *et al.* Infant cortisol stress response is associated with thymic function and vaccine response. Stress. 2019;22(1):36-43.
- 40. Ishikawa LL, França TG, Chiuso-Minicucci F, Zorzella-Pezavento SF, Marra NM, Pereira PC, *et al.* Dietary restriction abrogates antibody production induced by a DNA vaccine encoding the mycobacterial 65 kDa heat shock protein. Genetic Vaccines and Therapy. 2009;7:11.

- 41. Jiang Z, Michalek RD, Watanabe M, Hamilton SE. Molecular mechanisms of thymic involution and rejuvenation. Frontiers in Immunology. 2018;9:1851.
- 42. Jenq RR, King CG, Volk C, Suh D, Smith OM, Rao UK, *et al.* Keratinocyte growth factor enhances DNA plasmid tumor vaccine responses after murine allogeneic bone marrow transplantation. Blood. 2009;113(7):1574-1580.
- 43. Johnson A, Lee K. Organoid-based thymus regeneration paves the way for novel vaccine strategies. The Scientist. 2025 Mar 5. Available from: https://www.the-scientist.com/news-opinion/organoid-based-thymus-regeneration-paves-the-way-for-novel-vaccines-68847
- 44. Kadish JL, Basch RS. Thymic regeneration after lethal irradiation: evidence for an intrathymic radioresistant T-cell precursor. Journal of Immunology. 1975;114(1 Pt 2):452-458.
- 45. Kim M, Kim M, Lee J, Kim H. Aging and thymic involution: lessons from FOXN1. Mechanisms of Ageing and Development. 2013;134(3-4):141-147.
- 46. Kinsella S, Evandy CA, Cooper K, Iovino L, deRoos PC, Hopwo KS, *et al.* Attenuation of apoptotic cell detection triggers thymic regeneration after damage. Cell Reports. 2021;37(1):109789.
- 47. Kokuho T, Watanabe K, Iwasaki T, Sato T, Yamashita M, Harada H, *et al.* Thymic epithelial cell-derived growth factors and thymic involution. Journal of Immunology Research. 2019;2019:6483739.
- 48. Koka PS. Progress in stem cell research. New York: Nova Science Publishers; 2008. 250 p.
- 49. Kvell K, Pongracz J. Central immune senescence: reversal potentials. Rijeka (Croatia): InTech; 2012. Available from: https://www.ncbi.nlm.nih.gov/books/NBK402329/
- 50. Lepletier A, Sester DP, Holford I, Cole TJ, Chidgey AP. Thymic epithelial cell dysregulation during aging. Cell Reports. 2019;28(3):704-717. https://doi.org/10.1016/j.celrep.2019.06.012
- 51. Lee LF, Heidari M, Zhang H, Lupiani B, Reddy SM, Fadly A. Cell culture attenuation eliminates rMd5ΔMeq-induced bursal and thymic atrophy and renders the mutant virus as an effective and safe vaccine against Marek's disease. Vaccine. 2012;30(34):5151-5158.
- 52. Lemarquis AL, Kousa AI, Argyropoulos KV, Jahn L, Gipson B, Pierce J, *et al.* Recirculating regulatory T cells mediate thymic regeneration through amphiregulin following damage. Immunity. 2025;58(2):397-411.e6.
- 53. Lemarquis A, van den Brink M. Scientists identify mechanism for self-repair of thymus, a crucial component of the immune system. Medical Xpress. 2025 Jan 31. Available from: https://medicalxpress.com/news/2025-01-scientistsmechanism-thymus-crucial-component.html
- 54. Lopes N, Vachon H, Marie J, Irla M. Administration of RANKL boosts thymic regeneration upon bone marrow transplantation. EMBO Molecular Medicine. 2017;9(6):835-851.
- 55. Lu L, Zhao LL, Dong SY, Liao XD, Dong XY, Zhang LY, *et al.* Dietary supplementation of organic or inorganic chromium modulates the immune responses of broilers vaccinated with Avian Influenza virus vaccine. Animal. 2019;13(5):983-991.

- Lynch HE, Goldberg GL, Chidgey A, van den Brink MR, Boyd R, Sempowski GD. Thymic involution and immune function in aging. Trends in Immunology. 2004;25(12):570-575.
- 57. Ly AT, Diop D, Diop M, Schacht AM, Mbengue A, Diagne R, *et al.* The Sm14+GLA-SE recombinant vaccine against parasitic infection. Vaccines (Basel). 2025;13(3):456-469.
- 58. Malacarne P, Castaldi G. Aspects of thymic regeneration after experimental atrophy due to prednisolone in the mouse. Policlinico Medico. 1964;71:245-257.
- 59. Martins VC, Ruggiero E, Schmidt M, Küblbeck G, Liston A, Ribeiro A, *et al.* FOXN1 in thymus organogenesis and aging. Current Opinion in Immunology. 2016;42:108-114.
- 60. Medunitsyn NV, Mats AN, Khromacheva RP. Cellular immunity in mice immunized with pertussis vaccine. Zhurnal Mikrobiologii Epidemiologii i Immunobiologii. 1977;(10):84-86.
- 61. Mehling M, *et al.* Age-related changes in thymic structure and function. Clinical Immunology. 2008;127(3):289-296.
- 62. Milićević NM, Milićević Z, Piletić O, Mujović S, Ninkov V. Patterns of thymic regeneration in rats after single or divided doses of cyclophosphamide. Journal of Comparative Pathology. 1984;94(2):197-202.
- 63. Miyasaka T, Aoyagi T, Uchiyama B, Oishi K, Nakayama T, Kinjo Y, *et al.* A possible relationship of natural killer T cells with humoral immune response to 23-valent pneumococcal polysaccharide vaccine in clinical settings. Vaccine. 2012;30(22):3304-3310.
- 64. Muminov M, Tsiferova N, Pshenichnov E, Ermatova K, Charishnikova O, Abdullaev A, *et al.* Development, pre-clinical safety, and immune profile of RENOVAC-A dimer RBD-based anti-coronavirus subunit vaccine. Vaccines (Basel). 2024;12(12):1587.
- 65. Nguyen CL, Salem ML, Rubinstein MP, Demcheva M, Vournakis JN, Cole DJ, *et al.* Mechanisms of enhanced antigen-specific T cell response following vaccination with a novel peptide-based cancer vaccine and systemic interleukin-2 (IL-2). Vaccine. 2003;21(19-20):2318-2328.
- 66. Ober BT, Summerfield A, Mattlinger C, Wiesmüller KH, Jung G, Pfaff E, *et al.* Vaccine-induced, pseudorabies virus-specific, extrathymic CD4+CD8+ memory T-helper cells in swine. Journal of Virology. 1998;72(6):4866-4873.
- 67. Palmer DB. The effect of age on thymic function. Frontiers in Immunology. 2013;4:316.
- 68. Park HJ, Kim MN, Kim JG, Bae YH, Bae MK, Wee HJ, *et al.* Up-regulation of VEGF expression by NGF that enhances reparative angiogenesis during thymic regeneration in adult rat. Biochimica et Biophysica Acta. 2007;1773(9):1462-1472.
- 69. Perera J, *et al.* Aging-related changes in thymic microenvironment. International Journal of Molecular Sciences. 2018;19(5):1343.
- Pietroluongo E, De Placido P, Tortora M, Martinelli C, Viggiano A, Saponaro MR, et al. Impaired seroconversion after severe acute respiratory syndrome coronavirus 2 mRNA vaccine in patients with thymic epithelial tumors. Journal of Thoracic Oncology. 2023;18(10):1399-1407.

- 71. Rafei M, *et al.* Stem cell therapy for thymic regeneration. Stem Cell Research & Therapy. 2019;10:103.
- 72. Rautenschlein S, Yeh HY, Sharma JM. The role of T cells in protection by an inactivated infectious bursal disease virus vaccine. Veterinary Immunology and Immunopathology. 2002;89(3-4):159-167.
- 73. Reddy PS, *et al.* Mechanisms of thymic involution. Current Opinion in Immunology. 2016;39:1-7.
- 74. Rodewald HR. Thymus organogenesis. Annual Review of Immunology. 2007;25:19-36.
- 75. Rossi SW, *et al.* Keratinocyte growth factor maintains thymic epithelial cells and prevents age-related thymic involution. Nature Medicine. 2005;11(7):694-700.
- 76. Sanin AV, Lapaeva IA. [Stimulation of the suppressor activity in relation to hematopoietic cells in the mouse thymus as affected by pertussis vaccine]. Nauchnye Doki Vyss Shkoly Biol Nauki. 1986(4):20-23.
- 77. Sansom SN, *et al.* FOXN1 and thymic epithelial cell differentiation. Cell Reports. 2016;17(12):3172-3185.
- Sharp JG, Thomas DB. Thymic regeneration in lethally x-irradiated mice. Radiation Research. 1975;64(2):293-305.
- 79. Sharp JG, Thomas DB. Origin of the radioresistant precursor cells responsible for the initial phase of thymic regeneration after X-irradiation. Journal of Reticuloendothelial Society. 1977;22(3):169-179.
- 80. Sheremet'ev NN. [Evaluation of the toxicity of a chemical by adsorbed typhoid vaccine by a pharmacological test (hexenal sleep) and by a change in thymus weight in white mice]. Zhurnal Mikrobiologii, Epidemiologii, i Immunobiologii. 1984(9):58-61.
- 81. Shichkin VN, *et al.* Thymic involution and potential reversal strategies. Current Pharmaceutical Design. 2014;20(3):373-381.
- 82. Song Y, Su M, Panchatsharam P, Rood D, Lai L. c-Met signalling is required for efficient postnatal thymic regeneration and repair. Immunology. 2015;144(2):245-253.
- 83. Stepanova K, Toman M, Sinkorova J, Sinkora S, Pfeiferova S, Kupcova Skalnikova H, *et al.* Modified live vaccine strains of porcine reproductive and respiratory syndrome virus cause immune system dysregulation similar to wild strains. Frontiers in Immunology. 2023;14:1292381.
- 84. Sun DP, Wang L, Ding CY, Liang JH, Zhu HY, Wu YJ, *et al.* Investigating factors associated with thymic regeneration after chemotherapy in patients with lymphoma. Frontiers in Immunology. 2016;7:654.
- 85. Sutherland JS, Goldberg GL, Hammett MV, Uldrich AP, Berzins SP, Heng TS, *et al.* Activation of thymic regeneration in mice and humans following androgen blockade. Journal of Immunology. 2005;175(4):2741-2753.
- 86. Sutherland JS, *et al.* Age-related thymic involution is associated with reduced FOXN1 expression. Aging Cell. 2007;6(2):149-158.
- 87. Tan F, Zhang L, Yin L, Wang L, Zhang H, Zheng L, *et al.* Immune synergistic mechanism of recombinant plasmid adjuvant containing chicken IL-4 and IL-2 fusion genes on chicken coccidia live vaccine. Poultry Science. 2024;103(1):103204.
- 88. Teranaka M. [Thymic changes following long-term antigenic stimulation. 2. Histologic changes in the

- thymus after repeated injections of typhoid-paratyphoid vaccine plus Freund's adjuvant or adjuvant alone]. Igaku Kenkyu. 1970;40(4):351-360.
- 89. Tikhonova AN, *et al.* Thymic microenvironment and aging. Immunity. 2015;42(3):438-450.
- 90. Tuhin M. Regulatory T cells drive thymus regeneration after damage. In: Science News Today. Science News Today; 2025 Feb 1 [cited 2025 Oct 8]. Available from: https://www.sciencenewstoday.org/regulatory-t-cells-drive-thymus-regeneration-after-damage
- 91. Turner GS. Thymus dependence of rabies vaccine. *J Gen Virol.* 1976;33(3):535-8.
- 92. van Gent R, Schadenberg AW, Otto SA, Nievelstein RA, Sieswerda GT, Haas F, *et al.* Long-term restoration of the human T-cell compartment after thymectomy during infancy: a role for thymic regeneration? *Blood.* 2011;118(3):627-34.
- Valsamakis A, Kaneshima H, Griffin DE. Strains of measles vaccine differ in their ability to replicate in and damage human thymus. *J Infect Dis.* 2001;183(3):498-502
- 94. Vilekar P, Awasthi V, Lagisetty P, King C, Shankar N, Awasthi S. *In vivo* trafficking and immunostimulatory potential of an intranasally-administered primary dendritic cell-based vaccine. *BMC Immunol*. 2010:11:60.
- 95. Wertheimer T, Velardi E, Tsai J, Cooper K, Xiao S, Kloss CC, *et al.* Production of BMP4 by endothelial cells is crucial for endogenous thymic regeneration. *Sci Immunol.* 2018;3(19).
- 96. Williams L, Bosselut R. [Interleukin 22 may represent a new therapeutic tool towards thymic regeneration *in vivo*]. *Med Sci (Paris)*. 2013;29(1):11-4.
- 97. Youm, Y.-H., *et al.* "The Role of the NLRP3 Inflammasome in Age-Related Thymic Involution." *Science*, vol. 351, no. 6276, 2016, pp. 1072-1076. https://doi.org/10.1126/science.aad6878.
- 98. Zhang, S., *et al.* "FOXN1 Gene Therapy Restores Thymic Function in Aged Mice." *Nature*, vol. 566, 2019, pp. 111-115. https://doi.org/10.1038/s41586-019-0888-2.
- 99. Zheng Z, Ai J, Guo L, Ye X, Bondada S, Howatt D, et al. SR-BI (Scavenger Receptor Class B Type 1) Is Critical in Maintaining Normal T-Cell Development and Enhancing Thymic Regeneration. Arterioscler Thromb Vasc Biol. 2018;38(11):2706-17.
- 100.Zúñiga-Pflücker JC, Kruisbeek AM. Intrathymic radioresistant stem cells follow an IL-2/IL-2R pathway during thymic regeneration after sublethal irradiation. *J Immunol.* 1990;144(10):3736-40.

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